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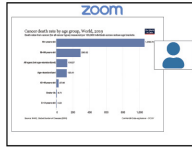
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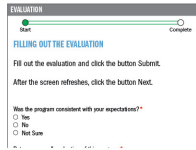
Zoom with Video to See the Slides

OR



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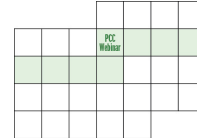
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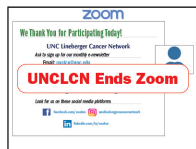


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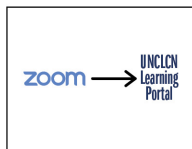
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After the Webinar

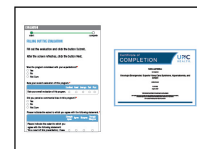
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May 21 - May 22



unclineberger.org/symposium

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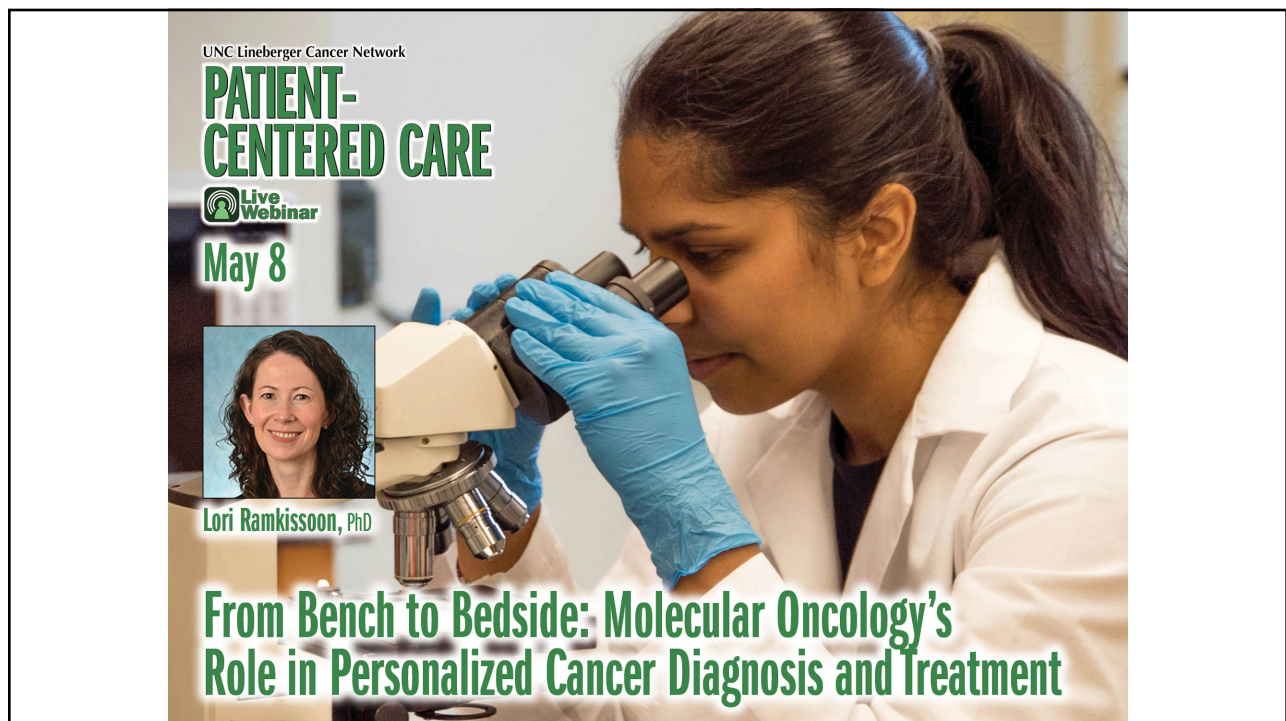
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UNC Lineberger Cancer Network

PATIENT-CENTERED CARE

 Live Webinar

May 8



Lori Ramkissoon, PhD

From Bench to Bedside: Molecular Oncology's Role in Personalized Cancer Diagnosis and Treatment

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Our Presenter



Lori Ramkissoon, PhD

Lori Ramkissoon, Ph.D., is a Clinical Assistant Professor of Pathology and Laboratory Medicine at the University of North Carolina Medical Center, where she leads the Cytogenetics Laboratory. This laboratory specializes in advanced genetic testing methodologies, including karyotyping, chromosomal microarray, and fluorescence in situ hybridization, to detect structural genetic variations in a broad spectrum of specimens ranging from prenatal and constitutional to oncological. These diagnostic services are crucial for identifying genetic disorders and pinpointing specific genetic markers that help classify various tumor types, thereby enhancing patient care through precise diagnoses.

Dr. Ramkissoon received a BA in Biochemistry from Baylor University and a Ph.D. from Weill Cornell Graduate School of Medical Sciences. Her postdoctoral tenure at the Dana-Farber Cancer Institute, under the mentorship of Dr. Keith Ligon, was pivotal in shaping her research focus on the genomic underpinnings of pediatric brain tumors. This experience motivated her to complete a clinical fellowship in Molecular and Clinical Cytogenetics at UNC, culminating in her board certification in Laboratory Genetics and Genomics. Additionally, Dr. Ramkissoon contributes her expertise to the UNC Precision Oncology Program, facilitating the incorporation of genomic insights into personalized treatment strategies for oncology patients.

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Our Presenter

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Our Presenter

4. Lori Ramkissoon, PhD is Director of the Cytogenetics laboratory at the University of North Carolina Medical Center

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Our Presenter

4. Lori Ramkissoon, PhD is Director of the Cytogenetics laboratory at the University of North Carolina Medical Center
3. She earned her PhD from Weill Cornell Graduate School of Medical Sciences and did a postdoctoral fellowship at Dana-Farber Cancer Institute

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Our Presenter

4. Lori Ramkissoon, PhD is Director of the Cytogenetics laboratory at the University of North Carolina Medical Center
3. She earned her PhD from Weill Cornell Graduate School of Medical Sciences and did a postdoctoral fellowship at Dana-Farber Cancer Institute
2. Prior to graduate school, she was a pre-doctoral fellow in the laboratory of Dr. Neal Young at the National Heart, Lung and Blood Institute in Bethesda, MD

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3. She earned her PhD from Weill Cornell Graduate School of Medical Sciences and did a postdoctoral fellowship at Dana-Farber Cancer Institute
2. Prior to graduate school, she was a pre-doctoral fellow in the laboratory of Dr. Neal Young at the National Heart, Lung and Blood Institute in Bethesda, MD
1. She worked for a year as a staff assistant in the United States Senate.

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Sample Poll Everywhere Question

Join by Web PollEv.com/unclcn Join by Text Send unclcn to 22333

Cancer is fundamentally a disease caused by changes to the "normal" sequence of a patient's genome, and the goal of molecular oncology is to define and understand these changes to benefit the diagnosis and treatment of cancer.

(A) True

(B) False

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ACCME Disclosure

This activity has been planned and implemented under the sole supervision of the Course Director, Stephanie Wheeler, PhD, MPH, in association with the UNC Office of Continuing Professional Development (CPD). The course director received research support from AstraZeneca (ended June 2023) and Pfizer Medical Foundation (ended December 2023). These financial relationships have been mitigated. CPD staff have no relevant financial relationships with ineligible companies as defined by the ACCME.

A potential conflict of interest occurs when an individual has an opportunity to affect educational content about health-care products or services of a commercial interest with which he/she has a financial relationship. The speakers and planners of this learning activity have not disclosed any relevant financial relationships with any commercial interests pertaining to this activity.

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ANCC Disclosure

NCPD Activity #: 001-L23079
1.0 Contact Hours Provided

Relevant Financial Relationship:

No one with the ability to control content of this activity has a relevant financial relationship with an ineligible company.

Criteria for Activity Completion:

Criteria for successful completion requires attendance at the NCPD activity and submission of an evaluation within 30 days.

Approved Provider Statement:

UNC Health is approved as a provider of nursing continuing professional development by the North Carolina Nurses Association, an accredited approver by the American Nurses Credentialing Center's Commission on Accreditation.

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Cancer is fundamentally a disease caused by changes to the "normal" sequence of a patient's genome, and the goal of molecular oncology is to define and understand these changes to benefit the diagnosis and treatment of cancer.

Response	Percentage
True	0%
False	0%

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From Bench to Bedside: Molecular Oncology's Role in Personalized Cancer Diagnosis and Treatment

Lori Ramkissoon, PhD

Lori.Ramkissoon@unchealth.unc.edu

May 8, 2024



SCHOOL OF
MEDICINE


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Learning Objectives

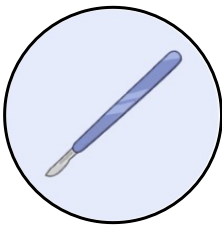
- Review advancements in the laboratory methods used to detect molecular biomarkers in oncology specimens
- Illustrate how molecular biomarkers have been integrated into diagnostic algorithms for certain cancer types
- Discuss the contributions of molecular oncology in treatment strategies



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Conventional treatment options for cancer patients





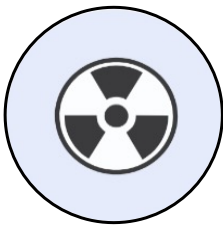
Surgery

- Resection
- Biopsy
- Fine needle aspirations




Chemotherapy

- Cytotoxic
- Neo-adjuvant or Adjuvant
- Maintenance regimens

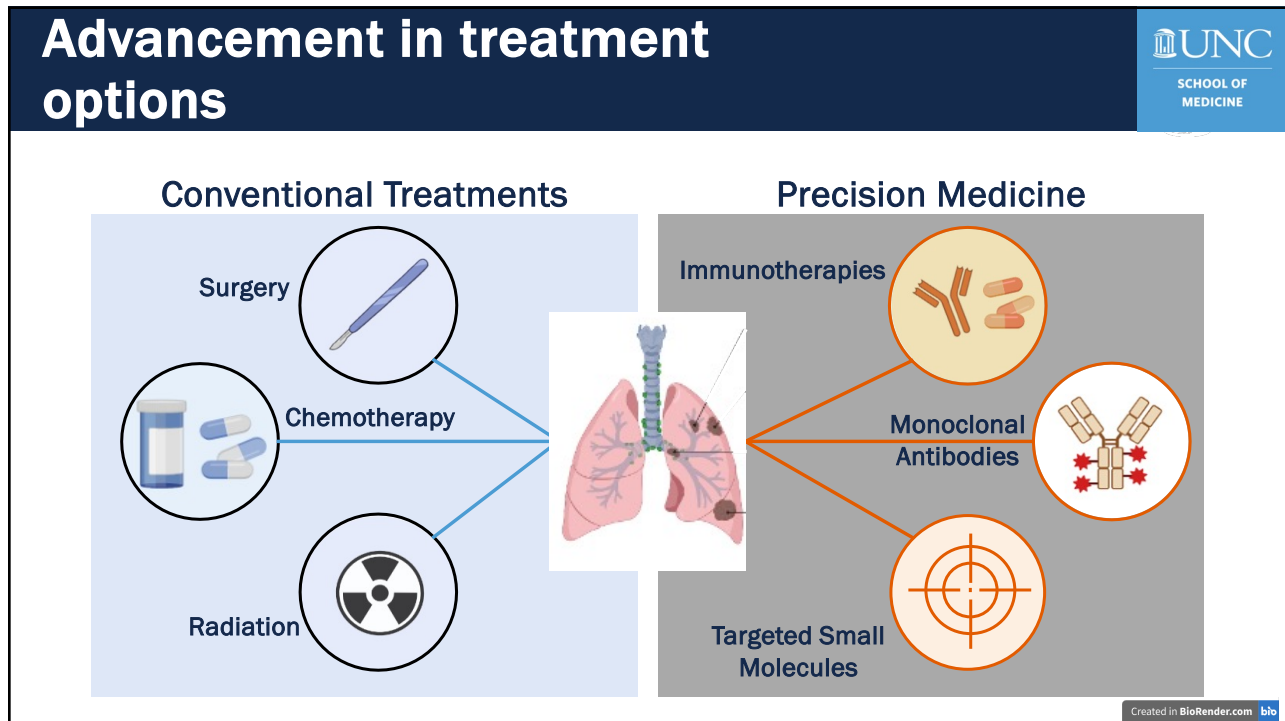


Radiation

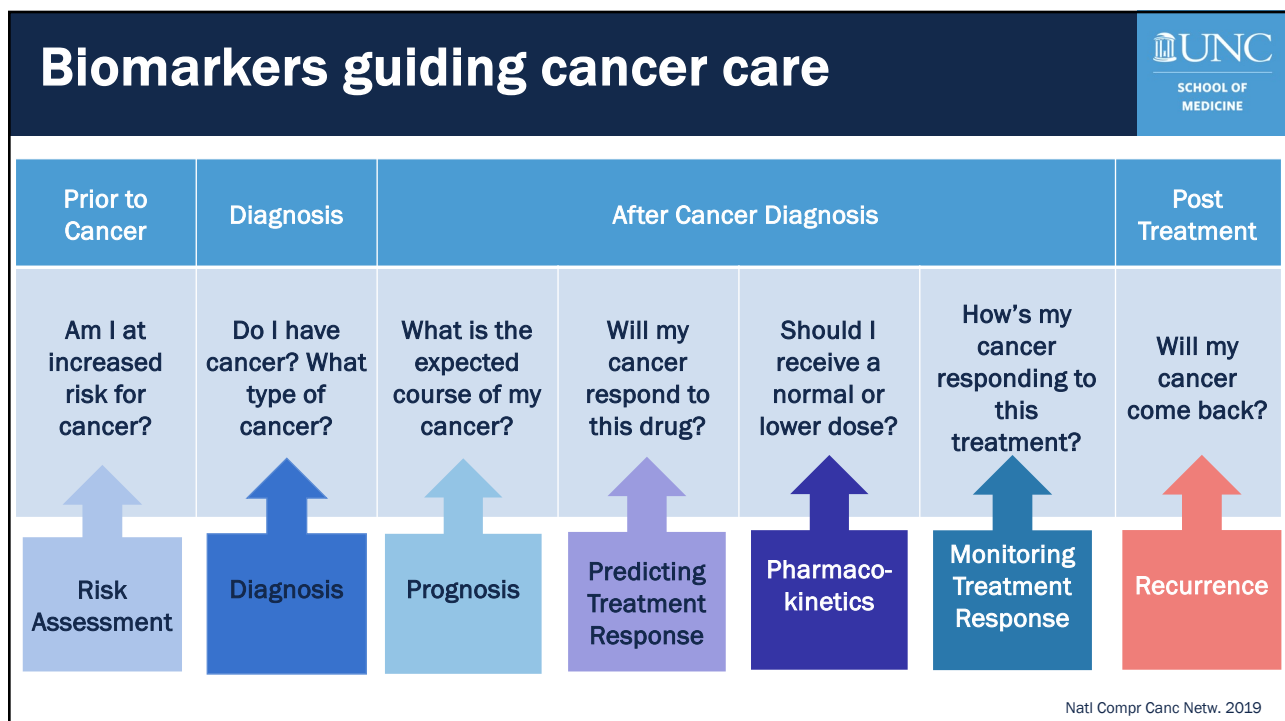
- External
- Internal

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Molecular biomarkers in cancer



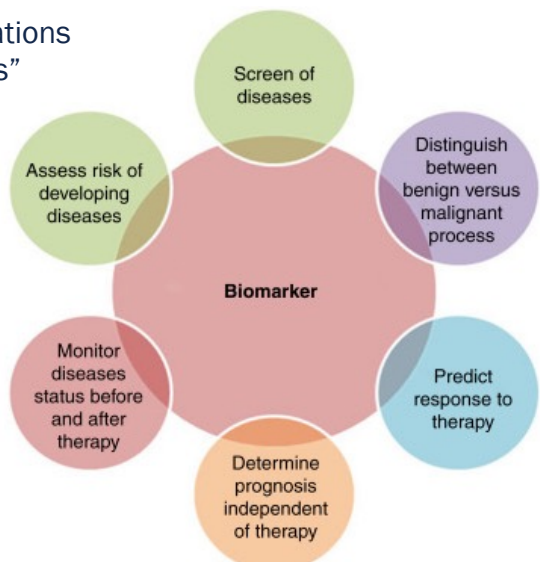
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What are biomarkers?



Biomarker: “cellular, biochemical or molecular alterations that are measurable in human tissues, cells, or fluids”

Biomarker	Example
Physiological biomarker	Blood pressure
Inflammatory biomarker	C-reactive protein
Prostate cancer biomarker	PSA
Molecular biomarker	<i>EGFR</i>
Somatic mutational biomarker	<i>KRAS G12D</i>
Germline mutational biomarker	<i>BRCA1</i>
Tumor agnostic biomarker	TMB, MSI, NTRK
Immune biomarker	PDL1



The diagram features a central pink circle labeled 'Biomarker'. Surrounding it are six other circles, each representing a clinical application: 'Screen of diseases' (green), 'Distinguish between benign versus malignant process' (purple), 'Predict response to therapy' (light blue), 'Determine prognosis independent of therapy' (orange), 'Monitor diseases status before and after therapy' (red), and 'Assess risk of developing diseases' (light green).

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How are molecular biomarkers used in oncology?



Diagnostic

- Assist with establishing diagnosis or classification
- Example: *BCR::ABL1* gene fusion in chronic myeloid leukemia (CML)

Prognostic

- Assist with determining the likely aggressiveness or course of disease
- Example: *TP53* mutations are an adverse prognostic factor in chronic lymphocytic leukemia (CLL)

Therapeutic

- Assist with prediction of response or resistance to a given drug, biologic, or regimen
- Example: *EGFR* activating mutations are associated with response to EGFR tyrosine kinase inhibitor (TKI) therapy in Non-Small Cell Lung Cancer

Clinical Trial Eligibility

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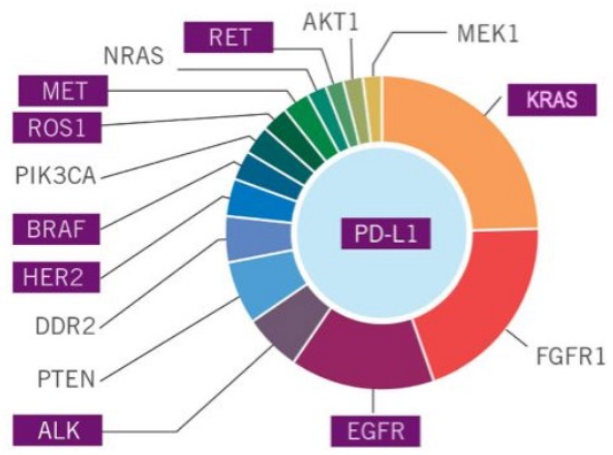
NCCN Guidelines and biomarkers



- Currently more than 800 biomarker recommendations are included in NCCN Guidelines
 - Determine risk of disease (BRCA-1/BRCA-2)
 - Screening (PSA for prostate)
 - Diagnostic (BCR/ABL in CML)
 - Prognostic (CA 19-9 in pancreas)
 - Predictive (ER/PR status in breast)
 - Risk of toxicity (UGT1A1*28 allele for irinotecan)
 - Response/disease monitoring (AFP; HCG in testicular)

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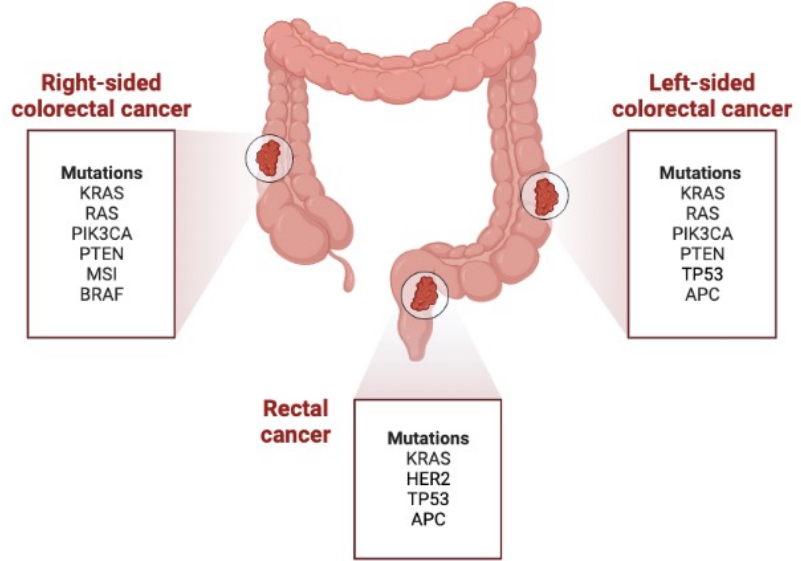
NCCN recommendations for biomarker testing in non-small cell lung cancer



Adapted from genetechnology.com and NCCN guidelines.

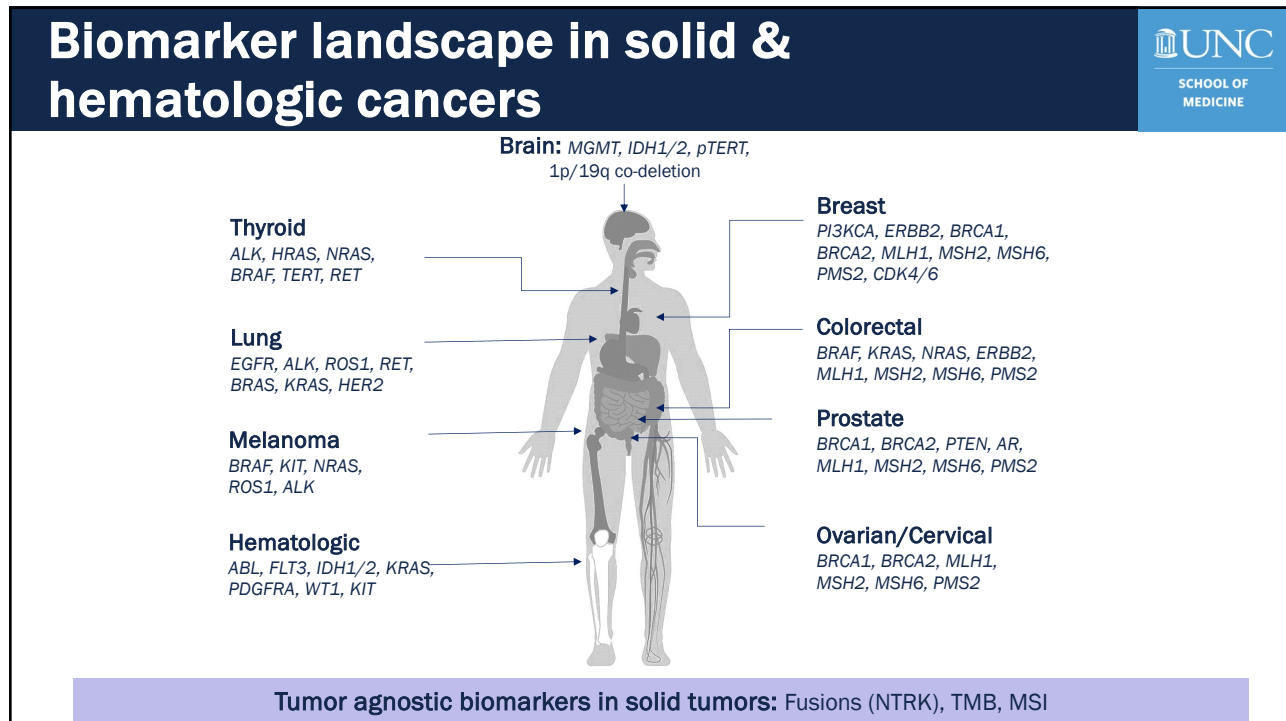
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NCCN biomarkers in colorectal cancer



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ASCO Guidelines

SOMATIC GENOMIC TESTING IN PATIENTS WITH METASTATIC OR ADVANCED CANCER

PROVISIONAL CLINICAL OPINION

WHICH METASTATIC OR ADVANCED SOLID TUMORS SHOULD UNDERGO GENOMIC SEQUENCING?

- Patients with metastatic or advanced solid tumors if there are genomic biomarker-linked therapies for that disease approved by the relevant regulatory agency (FDA)
- Patients with metastatic or advanced solid tumors if there are clearly defined resistance markers for a treatment being considered.

Chakravarty D et al. *J Clin Oncol* (2022) 40:1231-1258.

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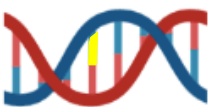
Detecting molecular biomarkers in cancer



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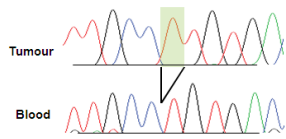
Types of genomic alterations that define cancer biomarkers





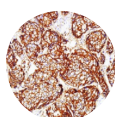
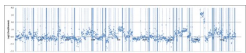
Base Pair Substitutions

- Limited to a single base pair/region within a single gene
- Examples: *EGFR* L858R, T790M; *BRAF* V600E, *IDH1* R132H



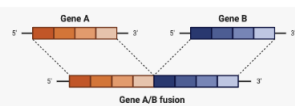
Insertions/deletions

- Limited to single genes and small changes in DNA sequence
- Examples: *EGFR* exon 19 deletions, *MET* exon 14

Copy Number Alterations

- Overexpression/amplification
- Examples: *HER2* amplification, *PDGFRA* amplification




Gene Rearrangements (Fusions)

- Detected via DNA and RNA (ASCO recommends RNA)
- Examples: *ALK* fusions, *NTRK* fusions

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Driver versus passenger alterations




Driver mutations (driving the bus)

- Mutations that give a cancer cell a competitive advantage
 - Increased rate of proliferation
 - Decreased apoptosis
 - Resistance to therapy
 - Etc.
- Contribute to oncogenesis (“oncogenic”)


Passenger mutations (along for the ride)

- Mutations that arise in cancer cells but don’t improve the “fitness” of the cancer cell
- Do not contribute to cancer development/progression



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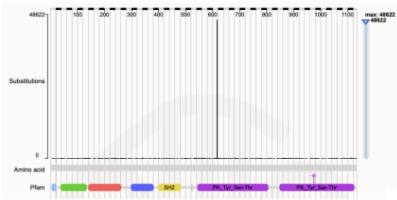
Gain-of-function versus Loss-of-function alterations



- Targetable mutations are typically GAIN OF FUNCTION mutations in oncogenes that encode for SIGNALING MOLECULES
 - GAIN OF FUNCTION mutations are frequently heterozygous (only one copy needs activated to drive the pathway)
 - There is generally a very limited number of mutations that can activate a protein
 - GAIN OF FUNCTION mutations tend to be recurrent among individuals

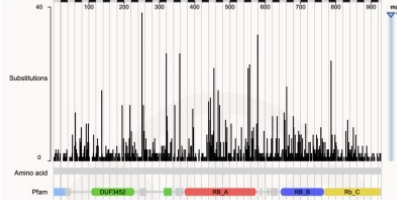
- LOSS OF FUNCTION mutations typically occur in TUMOR SUPPRESSOR GENES
 - Mutations tend to be LOSS OF FUNCTION, which requires BIALLELIC mutation
 - You need to lose BOTH copies to eliminate normal protein function
 - Because many different mutations can result in loss of function, there is typically a much broader spectrum of clinically significant mutations in tumor suppressor gene

JAK2




<https://cancer.sanger.ac.uk/cosmic/>

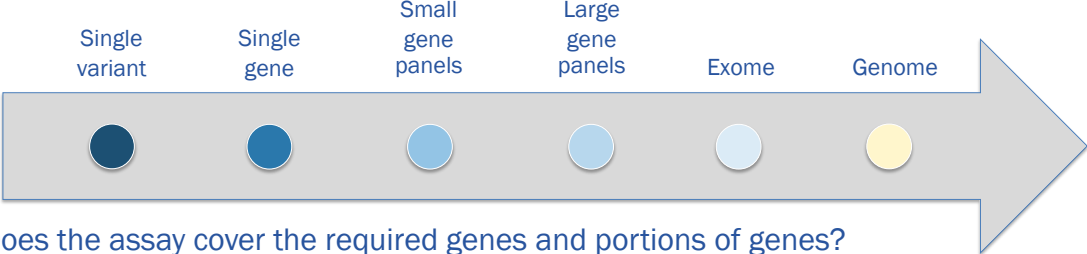
RB1



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Finding the right assay




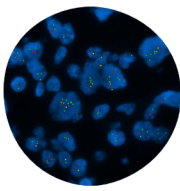


- Does the assay cover the required genes and portions of genes?
- What variant types can the assay detect?
 - Does the assay detect variant types required given the clinical indication for testing (single nucleotide variants, insertion-deletion, copy number gain/loss, gene rearrangements)?
- What are sample requirements?
- What is the expected turnaround time and cost?

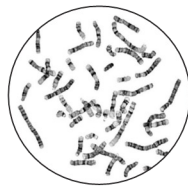
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Methodologies to detect cancer biomarkers

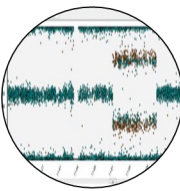




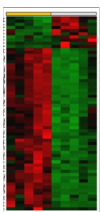
Fluorescence in situ hybridization (FISH)
Detect specific genomic translocations, copy number deletions/gains



Chromosome Analysis
Detects changes in chromosome number, translocations, large deletions/insertions



SNP microarrays
Used for genotyping and copy number determination



Gene expression arrays
Can provide subtype classification for certain tumors and/or prognostic information

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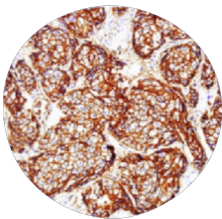
Methodologies to detect cancer biomarkers



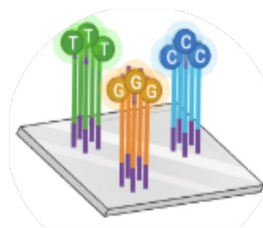
Single Gene Assays
Evaluate alterations in a single gene



Hotspot Panels
Sequencing of select hotspot codons, and not the entire coding region, of the genes included on the panel.



Immunohistochemistry
Determines protein expression within tissue sample



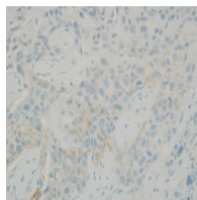
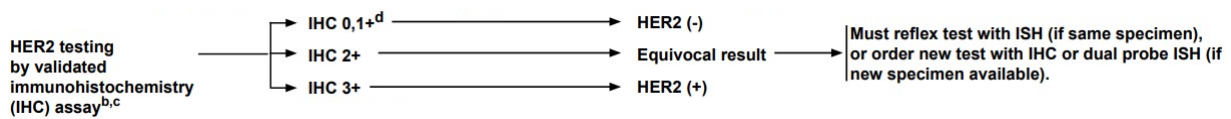
Broad Panel (Comprehensive Genomic Profiling)
An NGS test that sequences a defined list of genes with at least 50 genes in total. May also include RNA testing

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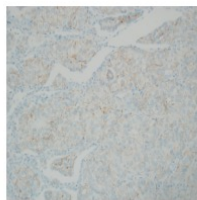
Methodologies are often combined to identify biomarkers



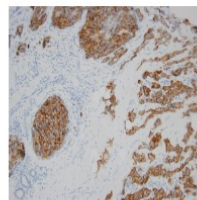
HER2 biomarker testing in invasive breast cancers



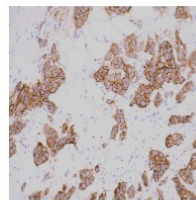
1+: IHC



2+: IHC



3+: IHC



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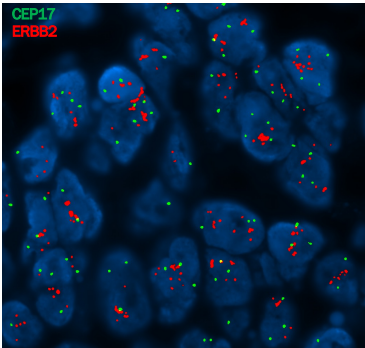
Methodologies are often combined to identify biomarkers

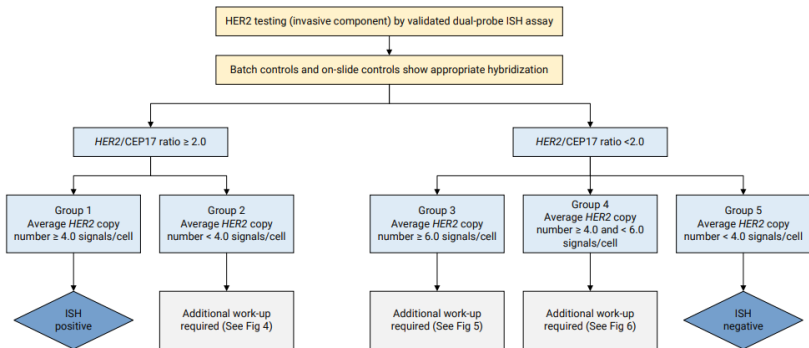
HER2 biomarker testing in invasive breast cancers

HER2 testing by validated immunohistochemistry (IHC) assay^{b,c}

- IHC 0,1+^d → HER2 (-)
- IHC 2+ → Equivocal result
- IHC 3+ → HER2 (+)

Must reflex test with ISH (if same specimen), or order new test with IHC or dual probe ISH (if new specimen available).





NCCN Guidelines Version 2.2024 Invasive Breast Cancer; Wolff AC et al. *J Clin Oncol* 2018;36:2105-2122

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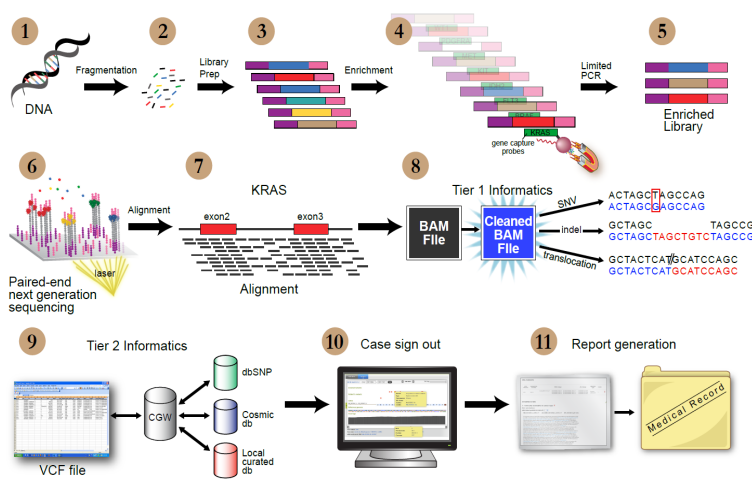
Overview of comprehensive genomic profiling

Goals

- High throughput, cost effective multiplexed sequencing assay with deep coverage
- Target clinically actionable regions important for all tumor types


Challenges

- Huge infrastructure costs
- Bioinformatic barriers
- Longer turnaround times



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Comparison of single gene testing versus comprehensive genomic profiling




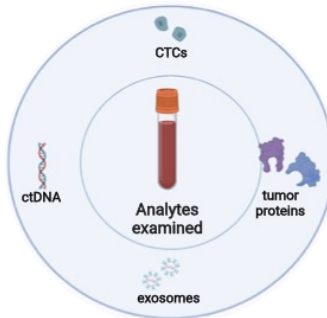
Single gene testing (SGT)	Comprehensive Genomic Profiling
<p>One or more tests ordered individually or simultaneously, but performed separately</p> <ul style="list-style-type: none"> PCR – defined regions for a limited number of mutations in <i>EGFR</i>, <i>KRAS</i> or <i>BRAF</i> FISH – known rearrangements in <i>ALK</i>, <i>RET</i>, <i>ROS1</i> or <i>MET</i> amplification IHC – known expression patterns or percentage of positive cells; loss of protein expression 	<p>One test ordered following negative SGT results or instead of SGT</p> <ul style="list-style-type: none"> DNA sequencing – simultaneously evaluate all major genomic variant types (mutations, copy number alterations, rearrangements) in oncogenes recommended for testing that have FDA-approved targeted therapies, as well those with emerging and potential clinical significance RNA sequencing – for known and novel rearrangements Microsatellite status and Tumor mutation burden – immunotherapy biomarkers

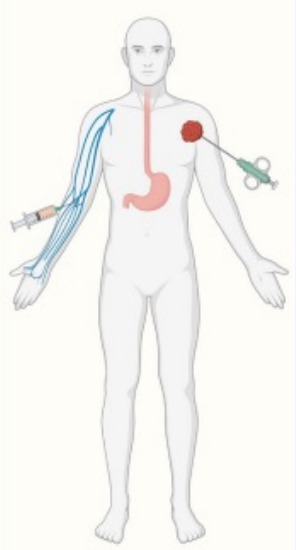
Nesline et al. *Oncol Ther* 2024 Mar 19.


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Cancer biomarker testing can be performed on a liquid or tissue biopsy








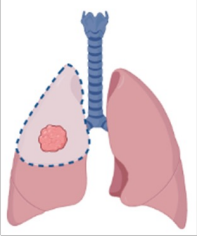


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Tissue acquisition: What material is available for testing?






Tissue Resection

- Obtained for diagnosis and symptomatic relief
- Tumor cell percentage may be an issue
- CGP and multiple assays typically not a problem for tumor-rich samples



Biopsy

- Obtained for a diagnosis
- Testing options may be limited but depends on tumor content not necessarily tissue size




Endobronchial Ultrasound (EBUS)/ Fine Needle Aspiration (FNA)

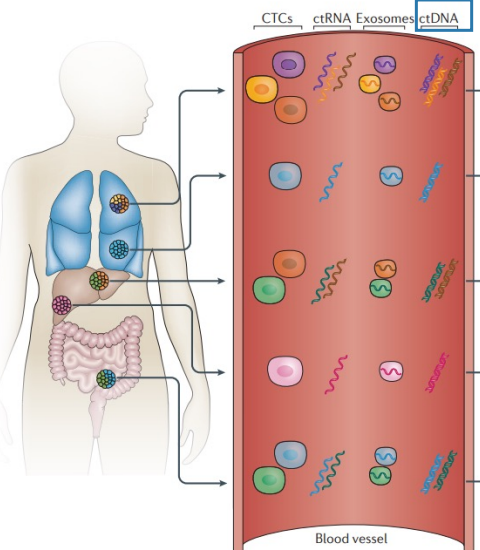
- Diagnosis can be made from very few cells
- Considered a cytology specimen
- May have significant limitations for testing

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Liquid biopsy: source of circulating tumor DNA (ctDNA)





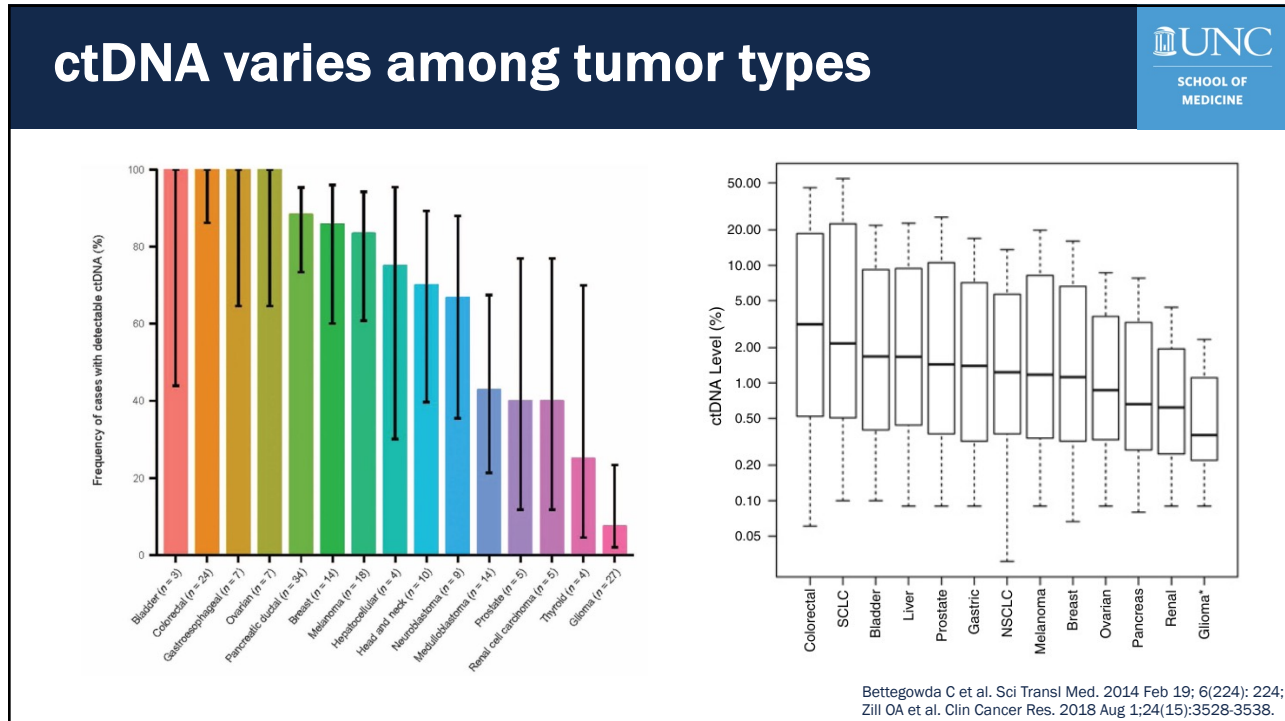
Blood vessel

- **ctDNA:** component of cell-free DNA which is tumor related
- **Cell-Free DNA Blood Collection Tubes:** specialized tubes required allow for isolation of plasma DNA up to 14 days after sample collection

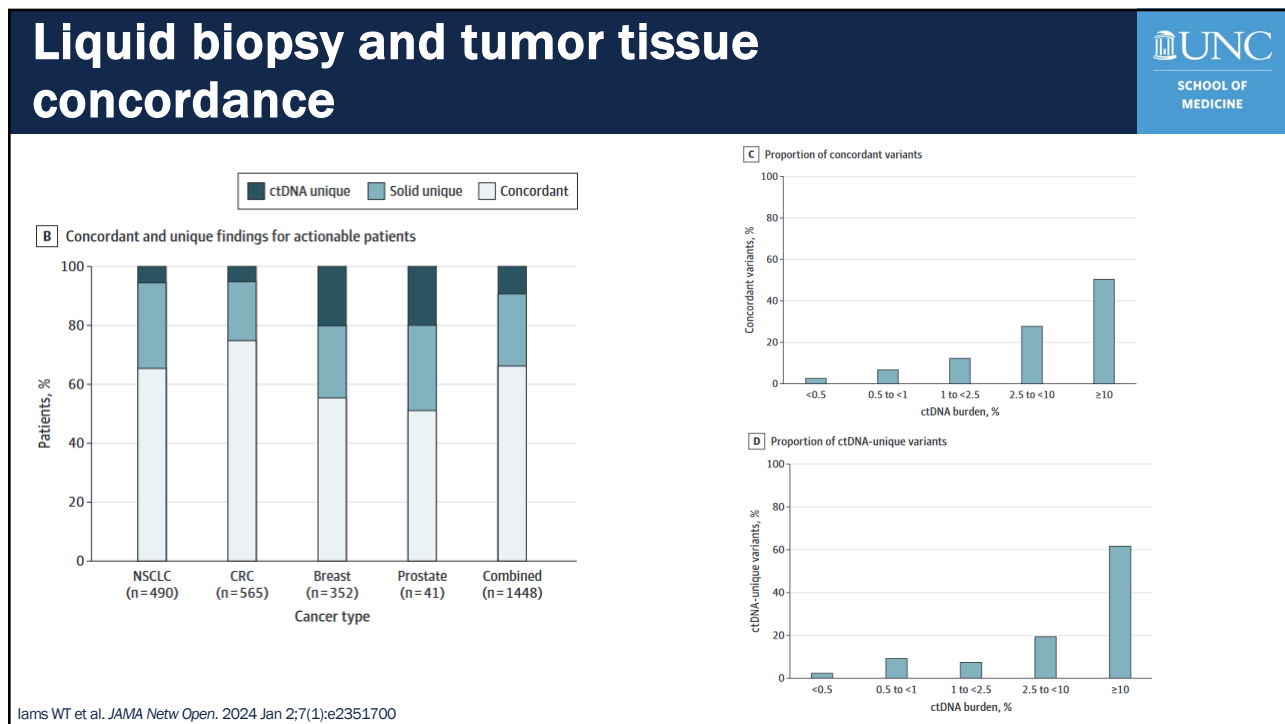


Nat Rev Clin Oncol. 2017 Sep;14(9):531-548

46



47




48

Advantages and disadvantages of tumor versus liquid biopsy

Tumor Biopsy

- Histological evaluation
- Tumor microenvironment analysis
- Clinical gold standard




- Surgery/needle biopsy
- Risk of complications
- Difficult to repeat & expensive
- Possible sampling bias
- Highly sensitive
- Longer TAT

- Blood draw
- Minimal complications
- Easy & repeatable
- Quick & cost-efficient
- Less sampling bias
- Rapid TAT
- False negatives
- Detection of CHIP

Liquid Biopsy

- Non-invasive
- Short half-life (<2 h)
- Compatible with longitudinal monitoring
- Representative of tumor heterogeneity





Adapted from Corcoran et al Nature Medicine 2020

49

ESMO Guidelines: Advanced cancer genotyping recommendations

Liquid Biopsy Best Practice

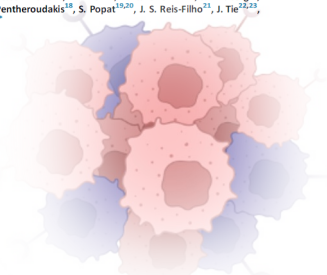
- ✓ May be used in clinical practice when results impact treatment.
 - ✓ May be used in clinical scenarios first where time to result is clinically important
 - ✓ Aggressive tumor type
 - ✓ No available tissue or biopsy not feasible
- ✓ Collect when tumor progressing (not regressing)
- ✓ Confirm testing if pathogenic variants of cancer susceptibility genes identified
- ✓ Negative tests should prompt tissue testing

SPECIAL ARTICLE

ESMO recommendations on the use of circulating tumour DNA assays for patients with cancer: a report from the ESMO Precision Medicine Working Group

J. Pascual¹, G. Attard², F.-C. Bidard^{3,4}, G. Curigliano^{5,6}, L. De Mattos-Arruda^{7,8}, M. Diehn⁹, A. Italiano^{10,11,12}, J. Lindberg¹³, J. D. Merker¹⁴, C. Montagut¹⁵, N. Normanno¹⁶, K. Pantel¹⁷, G. Pentheroudakis¹⁸, S. Popat^{19,20}, J. S. Reis-Filho²¹, J. Tie^{22,23}, J. Seoane^{24,25}, N. Tarazona^{26,27}, T. Yoshino²⁸ & N. C. Turner^{19,20*}



Adopted from Pascual et al. Annals of Oncology, 2022

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Poll everywhere question #2:



- Which testing methodology will most efficiently detect the recommended biomarkers (see table) in lung cancer?
 1. Individual FISH assays to interrogate each biomarker
 2. Hotspot DNA gene sequencing panel
 3. DNA and RNA sequencing assay

Recommended Testing
<i>ALK</i> rearrangements
<i>BRAF</i> mutations
<i>EGFR</i> mutations
<i>ERBB2</i> (HER2) mutations
<i>KRAS</i> mutations
<i>MET</i> exon 14 skipping mutations
<i>MET</i> amplification
<i>NTRK 1/2/3</i> rearrangements
<i>RET</i> rearrangements
<i>ROS1</i> rearrangements

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Which testing methodology will most efficiently detect the recommended biomarkers (see table) in lung cancer?

Individual FISH assays to interrogate each biomarker 0%

Hotspot DNA gene sequencing panel 0%

DNA and RNA sequencing assay 0%

Start the presentation to see live content. For screen share software, share the entire screen. Get help at pollev.com/app

52

Poll everywhere question #2: Answer

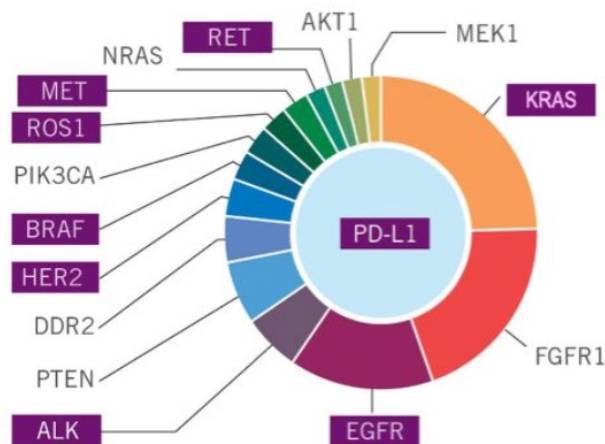


- Which testing methodology will most efficiently detect the recommended biomarkers (see table) in lung cancer?
 - Individual FISH assays to interrogate each biomarker
 - Hotspot DNA gene sequencing panel
 - DNA and RNA sequencing assay**

Recommended Testing
ALK rearrangements
BRAF mutations
EGFR mutations
ERBB2 (HER2) mutations
KRAS mutations
MET exon 14 skipping mutations
MET amplification
NTRK 1/2/3 rearrangements
RET rearrangements
ROS1 rearrangements

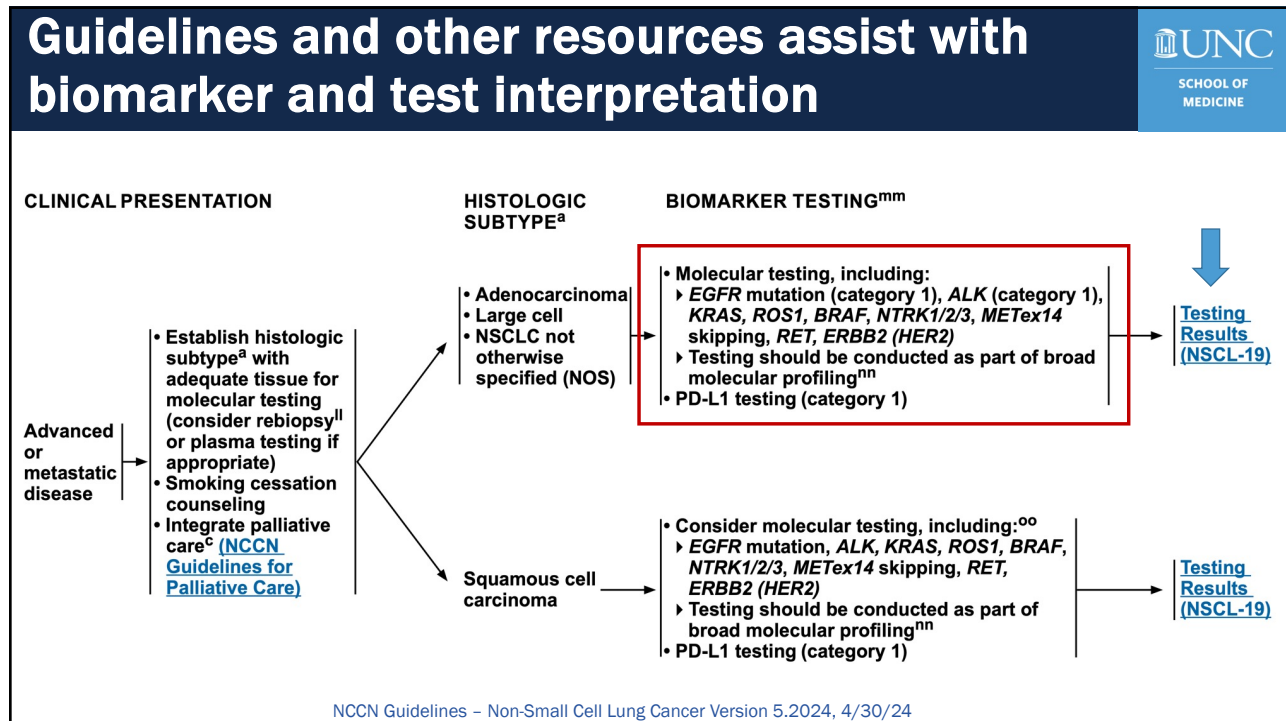
53

NCCN recommendations for biomarker testing in non-small cell lung cancer




Adapted from genetechnologycom and NCCN guidelines.

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55

Guidelines and other resources assist with biomarker and test interpretation

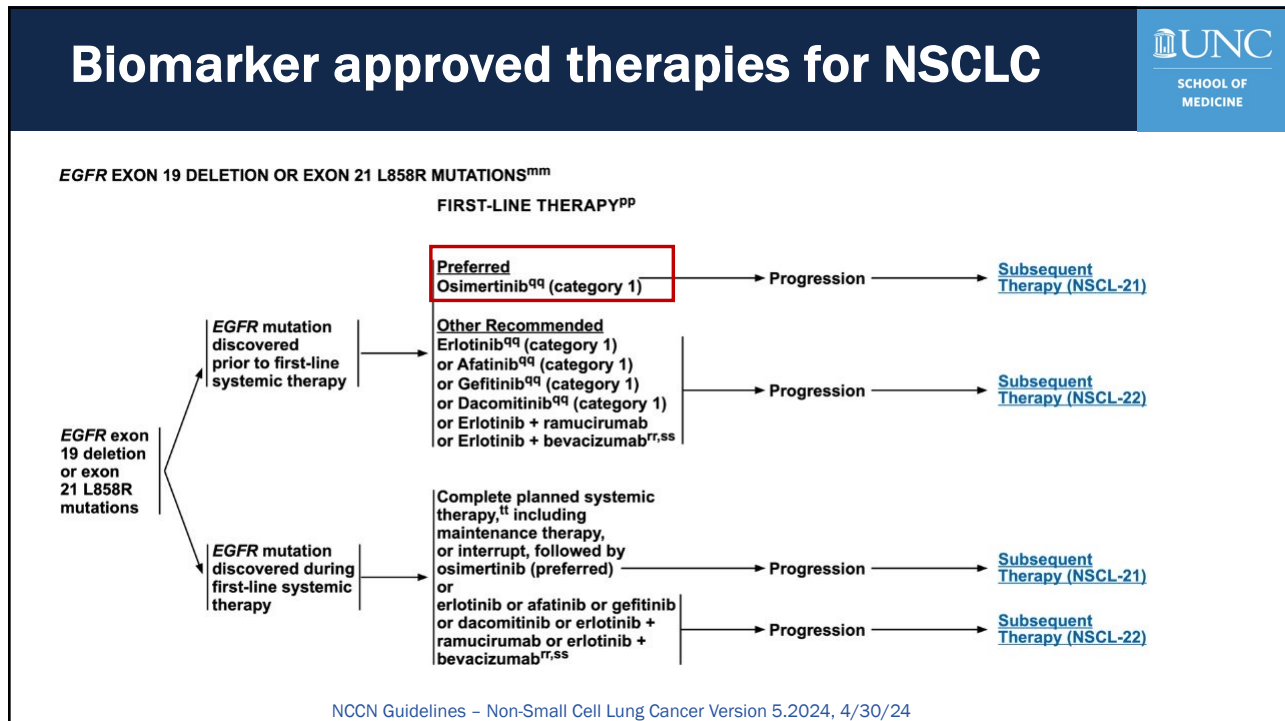


TESTING RESULTS^{ll,mm}

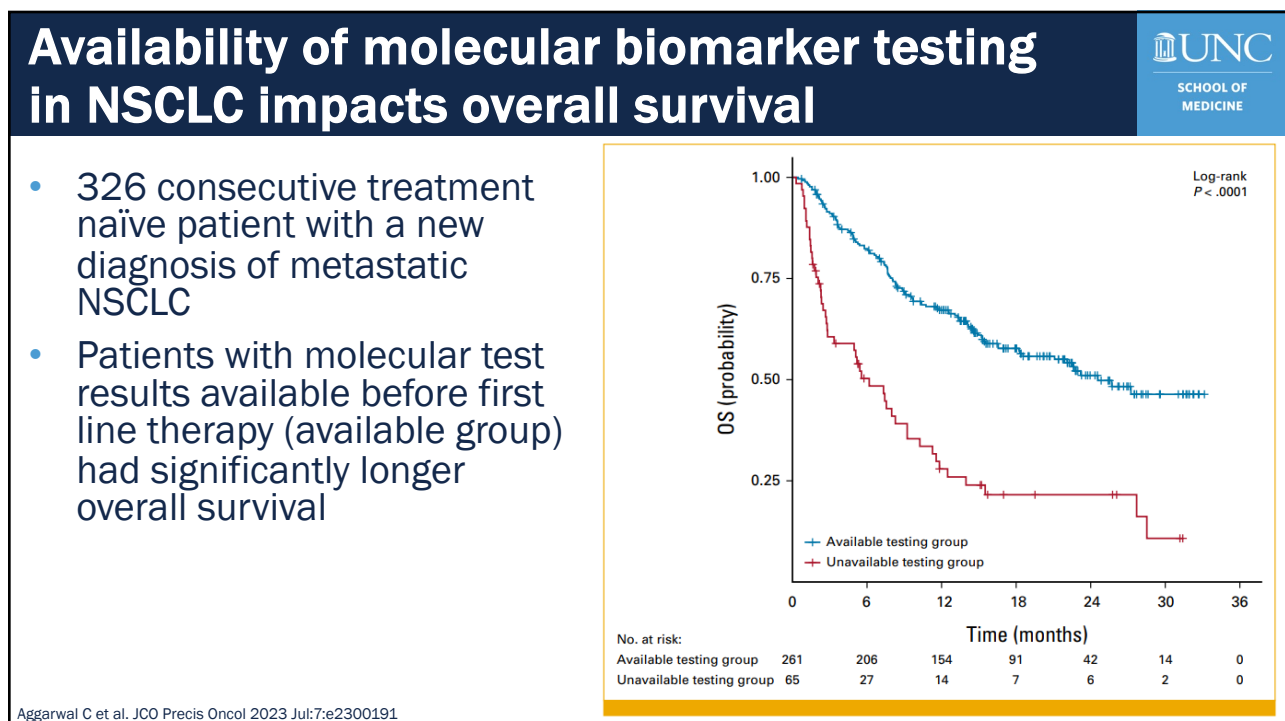
EGFR exon 19 deletion or exon 21 L858R mutation positive	NSCL-20 ←
EGFR S768I, L861Q, and/or G719X mutation positive	NSCL-23
EGFR exon 20 insertion mutation positive	NSCL-24
KRAS G12C mutation positive	NSCL-25
ALK rearrangement positive	NSCL-26
ROS1 rearrangement positive	NSCL-29
BRAF V600E mutation positive	NSCL-31
NTRK1/2/3 gene fusion positive	NSCL-32
METex14 skipping mutation positive	NSCL-33
RET rearrangement positive	NSCL-34
ERBB2 (HER2) mutation positive	NSCL-35
PD-L1 ≥1% and negative for actionable molecular biomarkers above	NSCL-36
PD-L1 <1% and negative for actionable molecular biomarkers above	NSCL-37

NCCN Guidelines – Non-Small Cell Lung Cancer Version 5.2024, 4/30/24

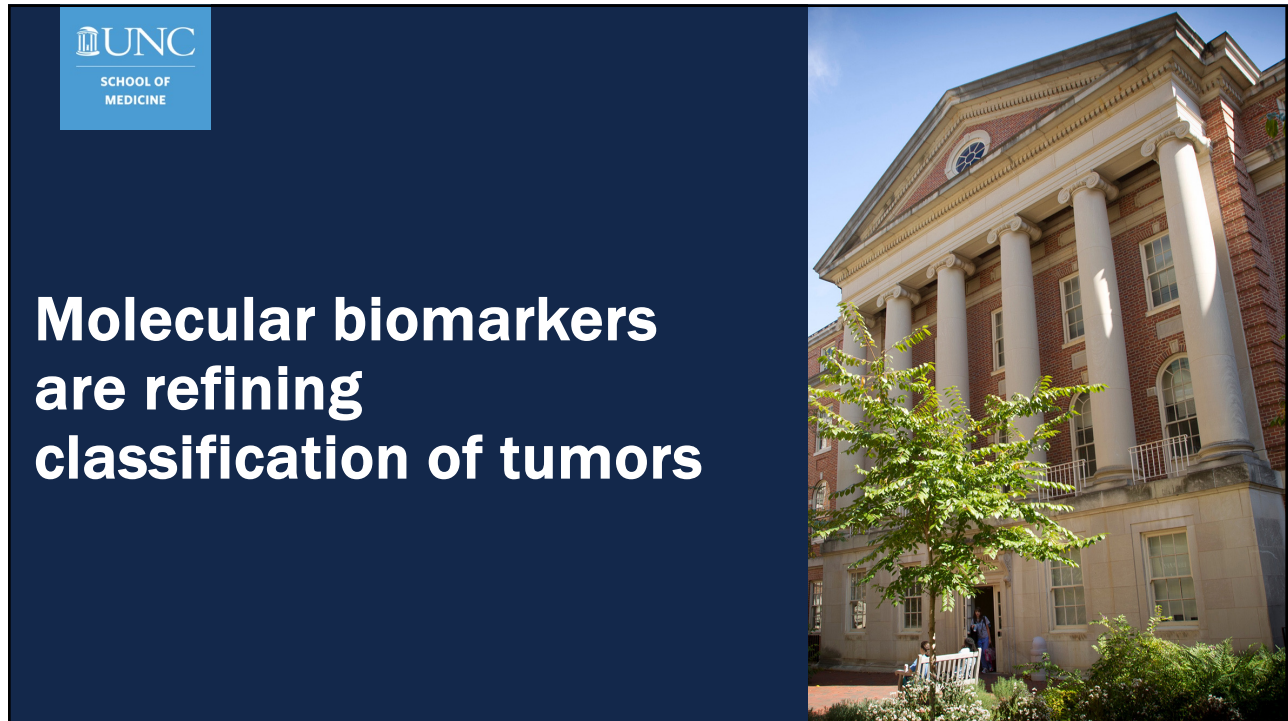
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Evolution of hematologic malignancy classification

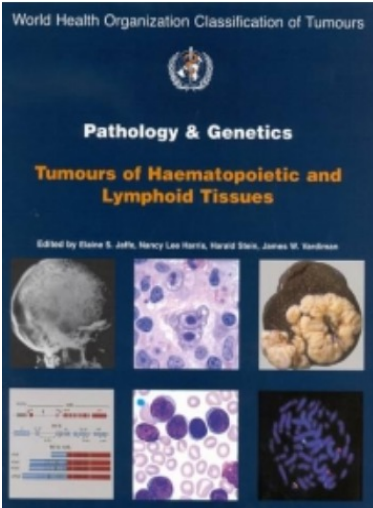
French-American-British (FAB) Classification System

FAB classification of acute myeloblastic leukaemia		FAB classification of acute myelomonocytic leukaemia	
<p>M0 Acute myeloblastic leukaemia with minimal differentiation</p> <p>Morphology: Can resemble L1/L2 blasts. Medium sized blasts, round nuclei, fine chromatin, basophilic non-granular cytoplasm, prominent nucleoli.</p> <p>Immunophenotype: -CD13 + -CD33 + -CD11b + -CD11c + -CD14 + -CD15 +</p>	<p>M4 Acute myelomonocytic leukaemia</p> <p>Morphology: Large blasts, moderate nucleocytoplasmic ratio and variable basophilia. The nucleus may be normal, kidney shaped or reniform. Nucleolus are usually prominent.</p> <p>Immunophenotype: -CD13 + -CD15 + -CD11b + -CD11c + -CD14 + -CD34 + -CD4 +</p>		
<p>M1 Acute myeloblastic leukaemia without maturation</p> <p>Morphology: Medium sized blasts with high nucleocytoplasmic ratio, rounded nuclei with irregular, dispersed chromatin with one or more prominent nucleoli. Blasts can show fine azurophilic granules or isolated Auer rods in the cytoplasm in 5% to 10% of cases.</p> <p>Immunophenotype: -MPO + -CD13 + -CD15 + -CD11b + -CD117 + -CD34 +/-</p>	<p>M5 Acute monocytic leukaemia</p> <p>Morphology: All acute monocytic leukaemia. Large blasts with rounded nucleus and abundant immature chromatin (1-3 nucleoli and moderately large and intensely specific cytoplasm. The cytoplasm may show some Auer rods and/or proproctopodia and progranulopodia.</p> <p>Immunophenotype: -CD14 + -CD11b + -CD11c + -HLA-DR + -CD4 +</p>		
<p>M2 Acute myeloblastic leukaemia with maturation</p> <p>Morphology: Small to medium-sized blasts with high nucleocytoplasmic ratio and rounded nuclei sometimes located in a corner of the cytoplasm. The nucleus shows dispersed, reniform chromatin with one or more nucleoli. The cytoplasm is basophilic and can contain traces of primary azurophilic granules or isolated Auer rods.</p> <p>Immunophenotype: -MPO + -CD13 + -CD15 + -CD11b + -HLA-DR +/- -Sudlow stain + -CD117 +/-</p>	<p>M6 Acute erythroid leukaemia</p> <p>Morphology: All erythroid leukaemias with proliferation of blast cells. Over 50% erythroid precursors and around 30% myeloblasts. The erythroid precursors are usually small, with thin, delicate, "spindled" or "multiform-shaped" cells, with azurophilic, "spindled" or "multiform-shaped" cells, and specialized azurophilic and proerythrocytic granules.</p> <p>Immunophenotype: -CD13 + -CD33 + -CD34 + -CD117 + -CD11c +/- -CD14 +/- -CD15 +/- -CD16 +/- -CD18 +/- -CD20 +/- -CD22 +/- -CD24 +/- -CD26 +/- -CD30 +/- -CD31 +/- -CD32 +/- -CD33 +/- -CD34 +/- -CD35 +/- -CD36 +/- -CD37 +/- -CD38 +/- -CD39 +/- -CD40 +/- -CD41 +/- -CD42 +/- -CD43 +/- -CD44 +/- -CD45 +/- -CD46 +/- -CD47 +/- -CD48 +/- -CD49 +/- -CD50 +/- -CD51 +/- -CD52 +/- -CD53 +/- -CD54 +/- -CD55 +/- -CD56 +/- -CD57 +/- -CD58 +/- -CD59 +/- -CD60 +/- -CD61 +/- -CD62 +/- -CD63 +/- -CD64 +/- -CD65 +/- -CD66 +/- -CD67 +/- -CD68 +/- -CD69 +/- -CD70 +/- -CD71 +/- -CD72 +/- -CD73 +/- -CD74 +/- -CD75 +/- -CD76 +/- -CD77 +/- -CD78 +/- -CD79 +/- -CD80 +/- -CD81 +/- -CD82 +/- -CD83 +/- -CD84 +/- -CD85 +/- -CD86 +/- -CD87 +/- -CD88 +/- -CD89 +/- -CD90 +/- -CD91 +/- -CD92 +/- -CD93 +/- -CD94 +/- -CD95 +/- -CD96 +/- -CD97 +/- -CD98 +/- -CD99 +/- -CD100 +/-</p>		
<p>M3 Promyelocytic leukaemia</p> <p>Morphology: Abundant, intensely azurophilic granulation. The nucleus is usually monocytic in appearance (reniform) and is either regular or bilobed with a deep cleft. Scarcely basophilic chromatin due to the proliferation of azurophilic granulation. Some atypical promyelocytes may contain elongated or spindle-shaped cytoplasmic inclusions (microgranules) specific to the type of leukaemia. These arise from storage, but differ from Auer rods in that they show a tubular substructure on electronic microscopy.</p> <p>Immunophenotype: -CD13 + -CD33 + -HLA-DR + -CD34 +</p>	<p>M7 Acute megakaryocytic leukaemia</p> <p>Morphology: Highly immature, proerythrocytic blasts. The nucleus is agranular with dispersed, irregularly dispersed and basophilic, and very dense in appearance to plasma, with occasional or granules. Microgranulopodia and fragments of megakaryocytic cells are seen in peripheral blood (seen in platelets, some highly megakaryocytic).</p> <p>Immunophenotype: -CD41 + -CD42 + -CD43 + -CD44 + -CD45 + -CD46 + -CD47 + -CD48 + -CD49 + -CD50 + -CD51 + -CD52 + -CD53 + -CD54 + -CD55 + -CD56 + -CD57 + -CD58 + -CD59 + -CD60 + -CD61 + -CD62 + -CD63 + -CD64 + -CD65 + -CD66 + -CD67 + -CD68 + -CD69 + -CD70 + -CD71 + -CD72 + -CD73 + -CD74 + -CD75 + -CD76 + -CD77 + -CD78 + -CD79 + -CD80 + -CD81 + -CD82 + -CD83 + -CD84 + -CD85 + -CD86 + -CD87 + -CD88 + -CD89 + -CD90 + -CD91 + -CD92 + -CD93 + -CD94 + -CD95 + -CD96 + -CD97 + -CD98 + -CD99 + -CD100 +</p>		

Ladines-Castro W, Barragán-Ibañez G, Luna-Pérez MA, et al. Morphology of leukaemias. Rev Médica del Hosp Gen México. 2016;79(2):107-113. doi:https://doi.org/10.1016/j.hgmx.2015.06.007

60

Evolution of hematologic malignancy classification



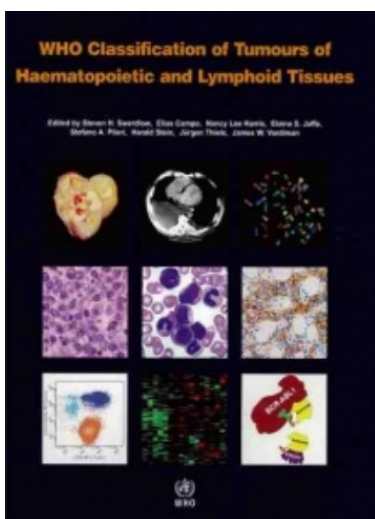
2001

I. Acute myeloid leukaemias with recurrent cytogenetic translocations

- AML with t(8;21)(q22;q22),AML1(CBFa)/ETO
- Acute promyelocytic leukaemia (AML with t(15;17)(q22;q11-12) and variants, PML/RARa)
- AML with abnormal bone marrow eosinophils (inv(16)(p13q22) or t(16;16)(p13;q11), CBFb/MYH11X)
- AML with 11q23 (MLL) abnormalities.

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Evolution of hematologic malignancy classification



2008

I. Acute myeloid leukemia with recurrent genetic abnormalities

- AML with t(8;21)(q22;q22); RUNX1-RUNX1T1
- AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22); CBFb-MYH11
- APL with t(15;17)(q22;q12); PML-RARA
- AML with t(9;11)(p22;q23); MLLT3-MLL
- AML with t(6;9)(p23;q34); DEK-NUP214
- AML with inv(3)(q21q26.2) or t(3;3)(q21;q26.2); RPN1-EV11
- AML (megakaryoblastic) with t(1;22)(p13;q13); RBM15-MKL1

II. Provisional entities:

- AML with mutated NPM1
- AML with mutated CEBPA

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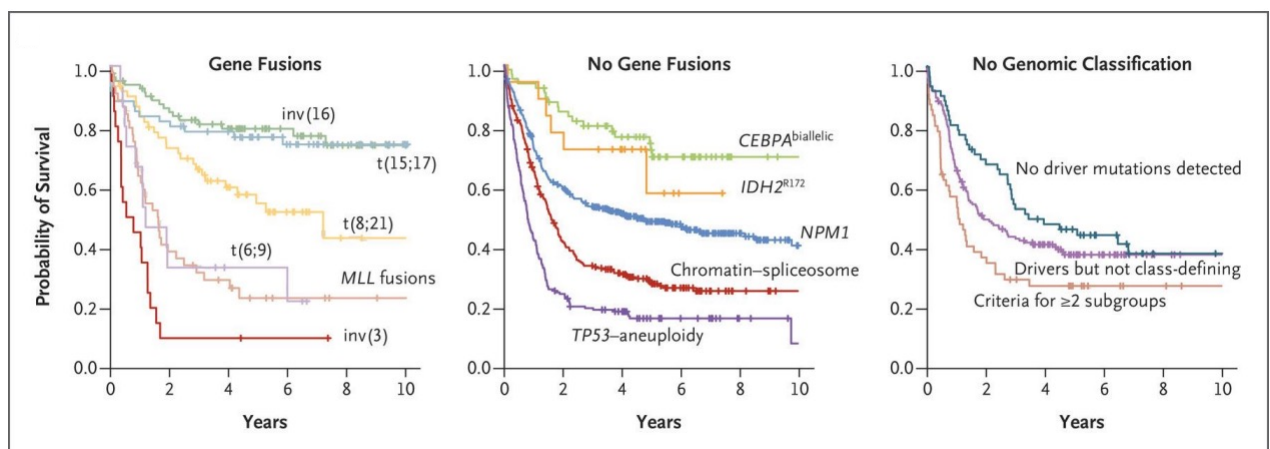
Evolution of hematologic malignancy classification



WHO 2017	WHO 2022	ICC 2022
AML with recurrent genetic abnormalities	AML with defining genetic abnormalities (no blast % cut-off, except*)	AML with recurrent genetic abnormalities (requiring equal or greater than 10% blasts, except *)
AML with t(8;21)(q22;q22.1);RUNX1-RUNX1T1	AML with RUNX1::RUNX1T1 fusion	AML with t(8;21)(q22;q22.1)/RUNX1::RUNX1T1
AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22);CBFB-MYH11	AML with CBFB::MYH11 fusion	AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22)/CBFB::MYH11
APL with PML-RARA	Acute promyelocytic leukaemia with PML::RARA fusion	Acute promyelocytic leukemia (APL) with t(15;17)(q24.1;q21.2)/PML::RARA; APL with other RARA rearrangements
AML with t(9;11)(p21.3;q23.3);MLLT3-KMT2A	AML with KMT2A rearrangement	AML with t(9;11)(p21.3;q23.3)/MLLT3::KMT2A; AML with other KMT2A rearrangements
AML with t(6;9)(p23;q34.1);DEK-NUP214	AML with DEK::NUP214 fusion	AML with t(6;9)(p22.3;q34.1)/DEK::NUP214
AML with inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2); GATA2-MECOM	AML with MECOM rearrangement	AML with inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2)/GATA2::MECOM(EVI1); AML with other MECOM rearrangements
AML (megakaryoblastic) with t(1;22)(p13.3;q13.3);RBM15-MKL1	AML with RBM15::MRTFA fusion	
Provisional entity: AML with BCR-ABL1	AML with BCR::ABL1 fusion*	AML with BCR::ABL1 fusion*
	AML with NUP98 rearrangement	
	AML with other (rare) defined genetic alterations*	AML with other rare recurring translocations

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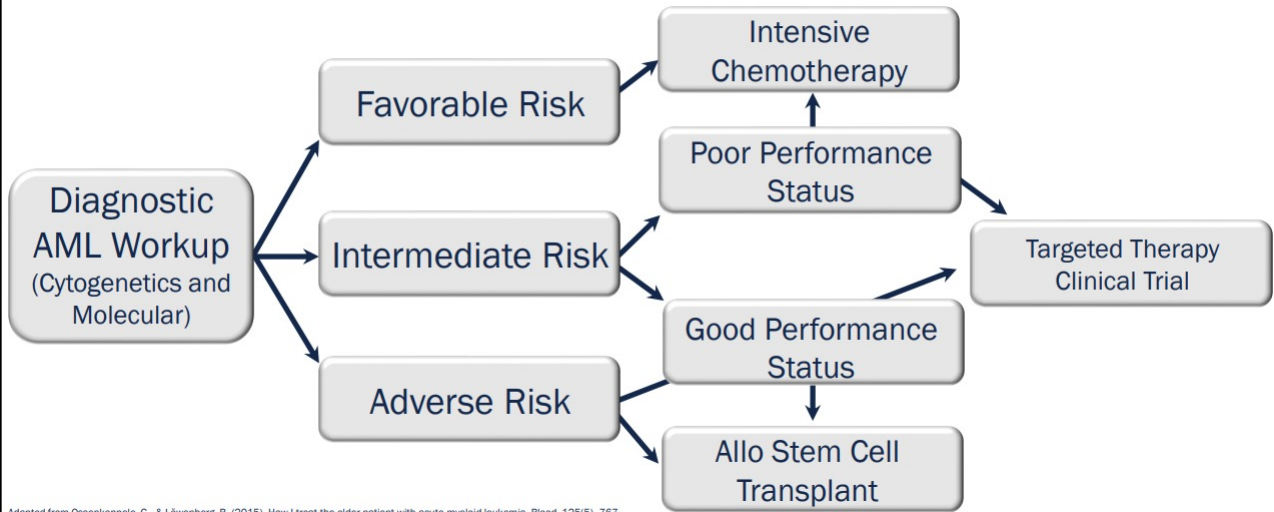
Impact of molecular testing in myeloid malignancy classification



Papaemmanuil E et al. N Engl J Med :374:2209-2221

64

Impact of molecular testing in myeloid malignancy classification



Adapted from Ossenkoppele, G., & Löwenberg, B. (2015). How I treat the older patient with acute myeloid leukemia. Blood, 125(5), 767.

65

Molecular biomarkers are increasingly incorporated into neuro-oncology



66

Evolution of molecularly classified CNS tumors



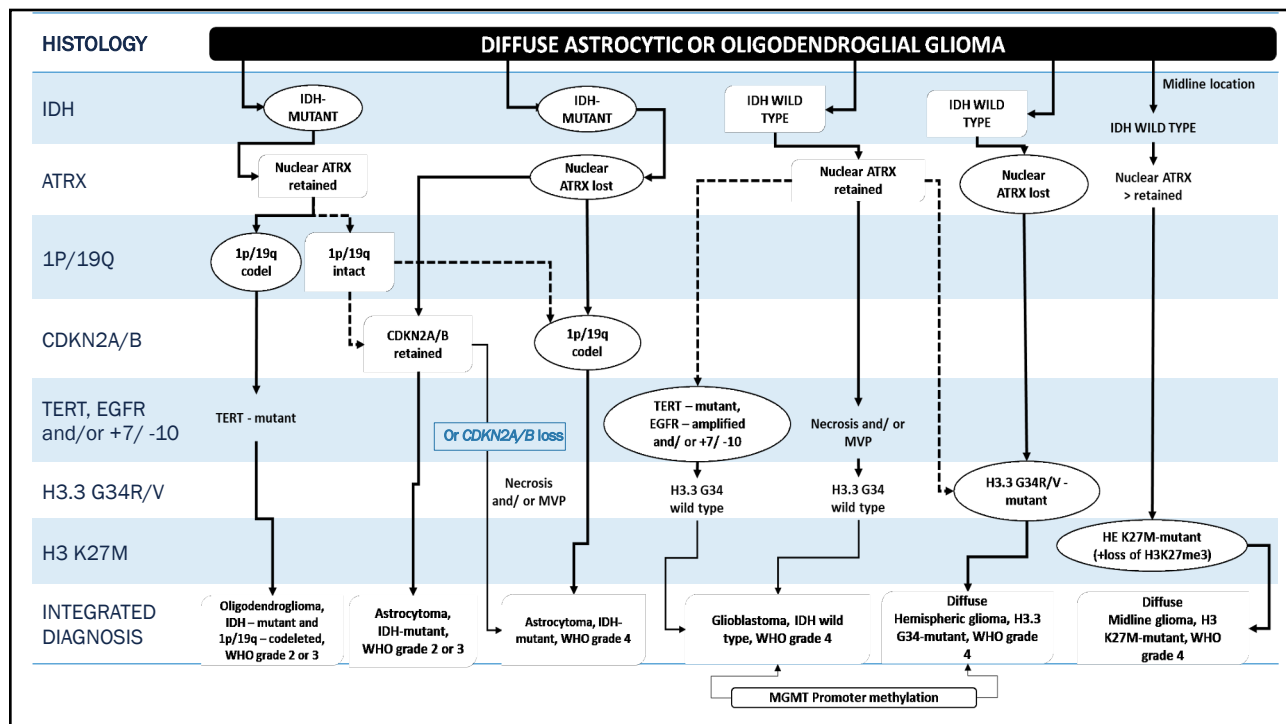
WHO 2016 GLIOMAS Diffuse Astrocytic and Oligodendroglial Tumours

Classification
Diffuse Astrocytoma, IDH-mutant
• Gemistocytic astrocytoma, IDH-mutant
Diffuse astrocytoma, IDH-wildtype
Diffuse astrocytoma, NOS
Anaplastic astrocytoma, IDH-mutant
Anaplastic astrocytoma, IDH – wildtype
Anaplastic astrocytoma, NOS
Glioblastoma, IDH-wildtype
• Giant cell glioblastoma
• Gliosarcoma
• Epithelioid glioblastoma
Glioblastoma, IDH-mutant
Glioblastoma, NOS
Diffuse midline glioma, H3 K27M mutant
Oligodendroglioma, IDH-mutant and 1p/19q-codeleted
Oligodendroglioma, NOS
Anaplastic, oligodendroglioma, IDH-mutant and 1p/19q-codeleted
Anaplastic oligodendroglioma, NOS
Oligoastrocytoma, NOS
Anaplastic oligoastrocytoma, NOS

WHO 2021 GLIOMAS Gliomas, Glioneural and Neuronal Tumours

- Adult-type diffuse gliomas**
 - Astrocytoma, IDH-mutant
 - Oligodendroglioma, IDH-mutant and 1p/19q-codeleted
 - Glioblastoma, IDH-wildtype
- Paediatric-type diffuse low-grade gliomas**
 - Diffuse astrocytoma, MYB or MYBL1 – altered
 - Angiocentric glioma
 - Polymorphous low-grade neuroepithelial tumour of the young
 - Diffuse low-grade glioma, MAPK pathway-altered.
- Paediatric-type diffuse high-grade gliomas**
 - Diffuse midline glioma, H3 K27- altered
 - Diffuse hemispheric glioma, H3 G34-mutant
 - Diffuse paediatric-type high-grade glioma, H3 wild-type and IDH wild type
 - Infant-type hemispheric glioma

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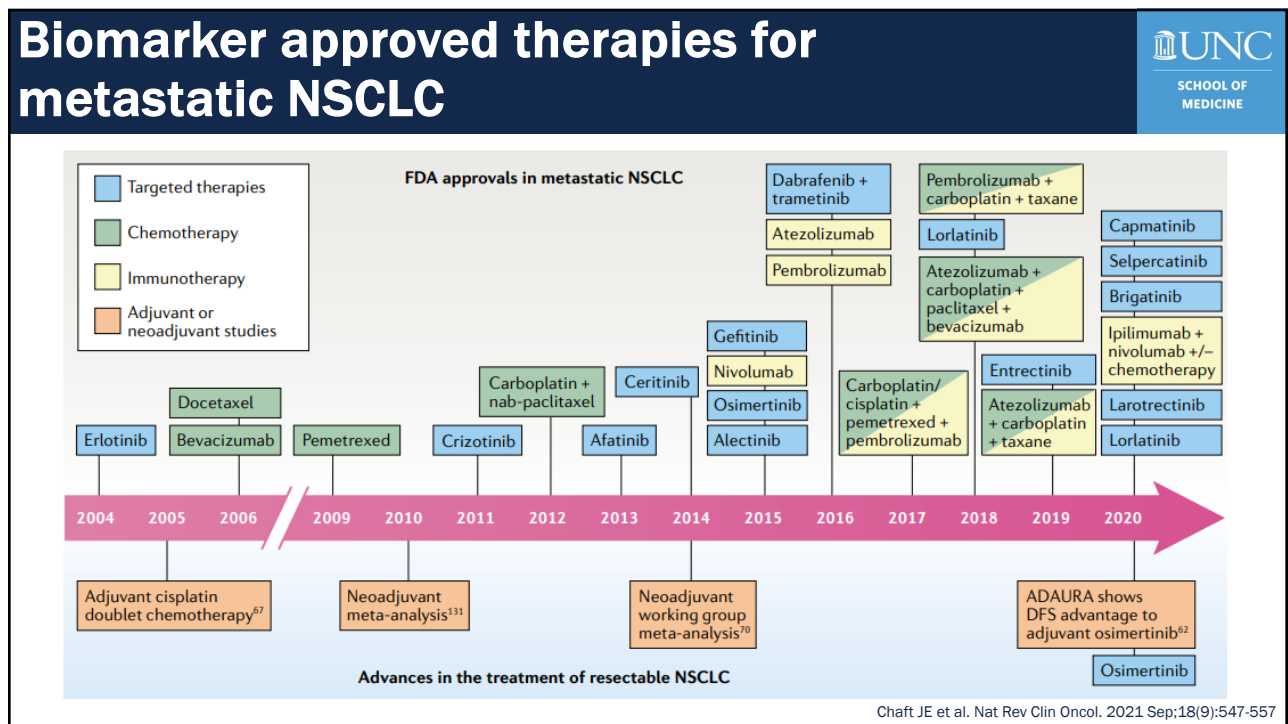


68

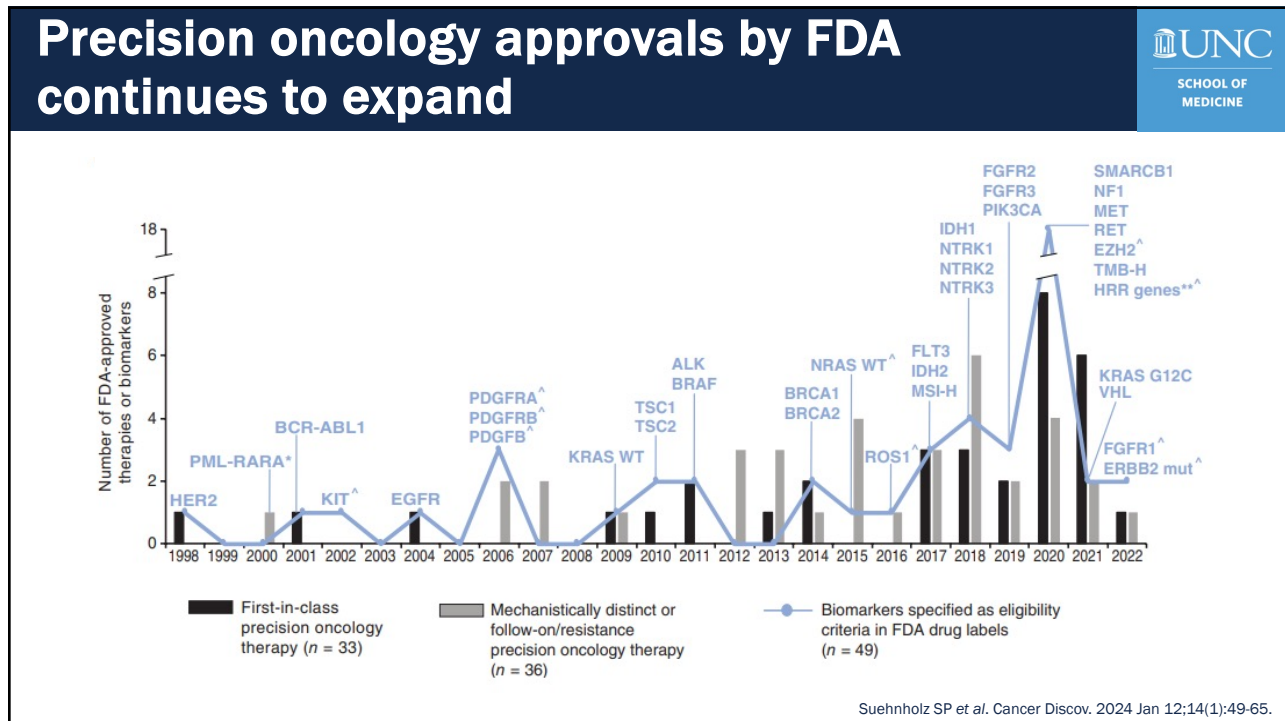


Molecular biomarkers guide treatment decisions

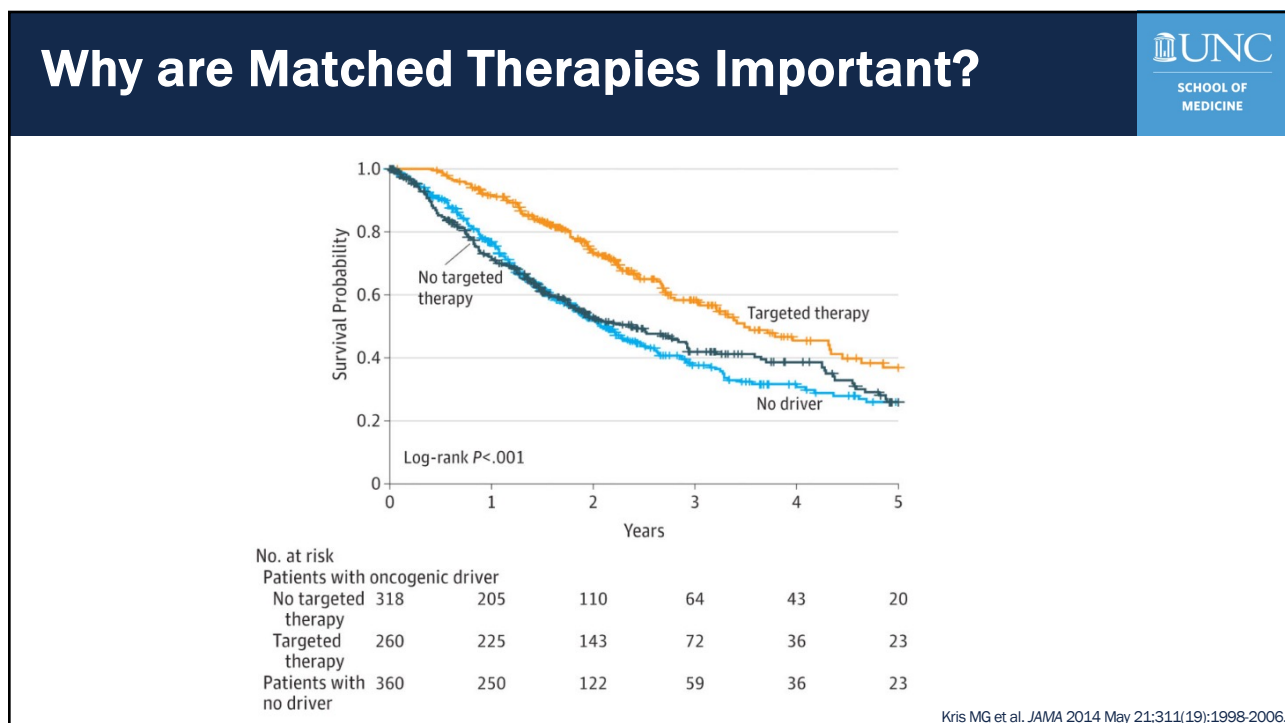
69



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72

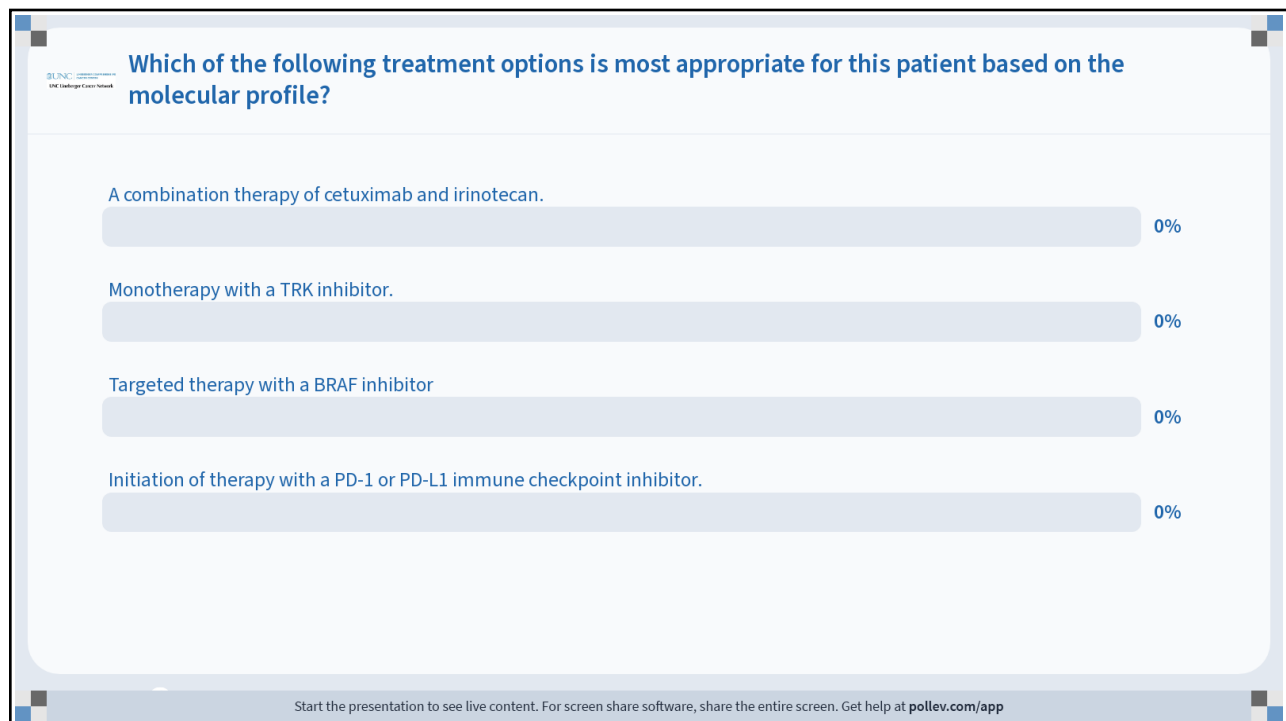
Poll everywhere question #3:



In a patient with newly diagnosed colon cancer, testing determined that the tumor had microsatellite instability (MSI) or was MSI-high. Which of the following treatment strategies is indicated for MSI-high status?

1. A combination therapy of cetuximab and irinotecan.
2. Monotherapy with a TRK inhibitor.
3. Targeted therapy with a BRAF inhibitor
4. Initiation of therapy with a PD-1 or PD-L1 immune checkpoint inhibitor.

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Poll everywhere question #3:

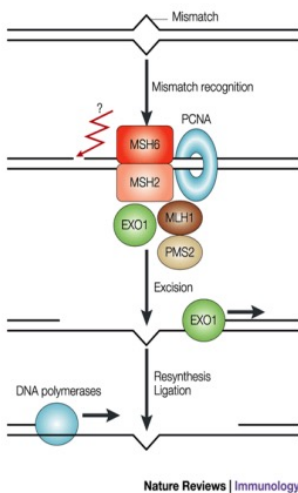


In a patient with newly diagnosed colon cancer, testing determined that the tumor had microsatellite instability (MSI) or was MSI-high. Which of the following treatment strategies is indicated for MSI-high status?

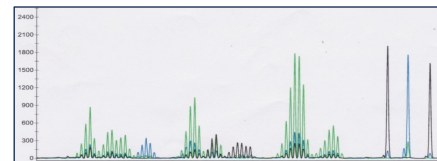
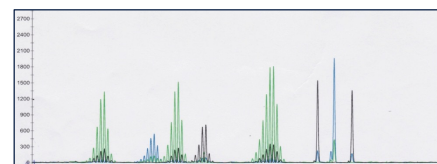
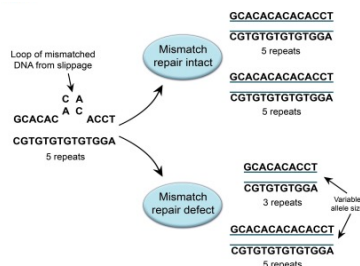
1. A combination therapy of cetuximab and irinotecan.
2. Monotherapy with a TRK inhibitor.
3. Targeted therapy with a BRAF inhibitor
4. **Initiation of therapy with a PD-1 or PD-L1 immune checkpoint inhibitor.**

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DNA Mismatch Repair and Microsatellite Instability



- During DNA replication, errors occur in microsatellite regions
 - Short repetitive DNA sequences (1-6 base pairs) or tandem repeats
- Multi-protein complex corrects these single base pair mismatches and small insertion-deletion errors
- Failures to repair these errors during replication leads to expansion of repeats and genomic instability = **microsatellite instability**

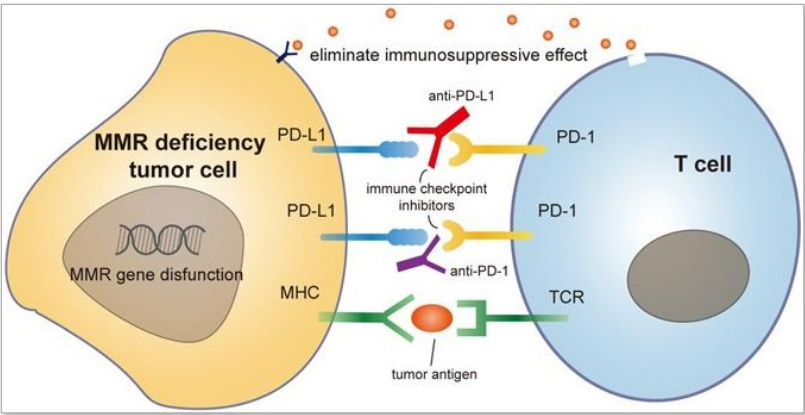


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Anti-PD-1/L1 therapies reactivate T cell activity



- MSI-High/dMMR tumors generate neoantigens which are recognized by the immune system
- Tumor cells also express PD-L1 to inhibit T-cell activity
- Immune checkpoint inhibitors reactivate T-cell activity



He Y et al. *Int J Biol Sci* 2022; 18(7):2821-2832

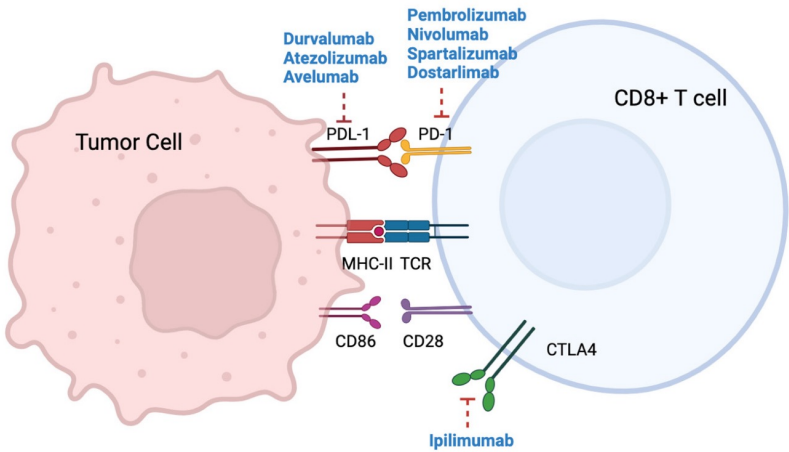
77

Tumor agnostic immune checkpoint inhibitor approval



FDA grants accelerated approval to pembrolizumab for first tissue/site agnostic indication


- 2017 approval for adult and pediatric patients with unresectable or metastatic solid tumors
 - microsatellite instability-high (MSI-H) or mismatch repair deficient (dMMR)
 - progressed following prior treatment

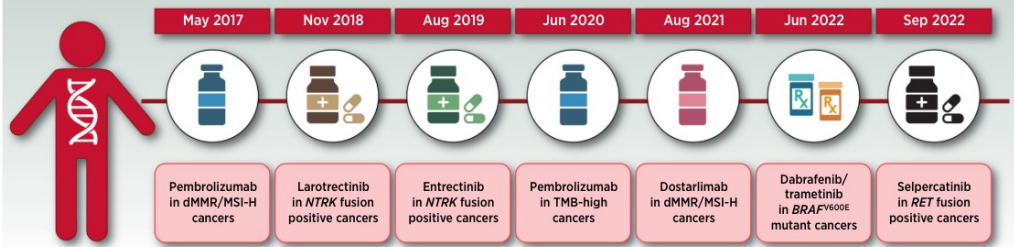


Ros J et al. *Cancers (Basel)*. 2023 Aug 24;15(17):4245.

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Additional FDA-approved tumor agnostic targeted therapies





- **NTRK1, NTRK2, NTRK3** transmembrane tyrosine kinases that are important for neuronal development
 - Fusions detected in 1.6% of profiled cases
 - *NTRK* fusions can be targeted using Larotrectinib and Entrectinib
- **Tumor mutation burden (TMB)** reflects the number of genetic alterations in the genome of cancer cells
 - Calculated using data from NGS of either tissue or plasma, mutations per megabase (mut/MB)
 - 10 mut/mb as for defining indication of pembrolizumab
- **BRAF V600E** can be targeted by using inhibitors of BRAF and MEK
 - Detected in 3% of AACR Project GENIE (version 13) pan-cancer cohort
 - Dabrafenib and Trametinib received FDA approval for BRAFV600E-positive solid tumors
- **RET fusion-positive** samples were identified in 1.5% of 25,972 tumors profiled for structural variants
 - Selpercatinib, ATP-dependent selective RET inhibitor, approved by FDA for solid tumors with *RET* fusions

Gouda MA et al. *Clin Cancer Res.* 2023 Aug 1;29(15):2753-2760

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Targeted therapy success story

Chronic Myeloid Leukemia



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Visualization of first chromosome translocation

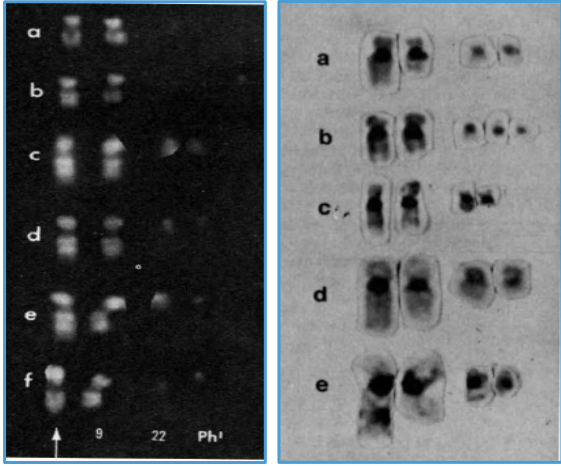


A New Consistent Chromosomal Abnormality in Chronic Myelogenous Leukaemia identified by Quinacrine Fluorescence and Giemsa Staining

JANET D. ROWLEY

Department of Medicine,
University of Chicago and
Franklin McLean Memorial Research Institute,
Chicago, Illinois 60637


Received January 8; revised February 8, 1973.



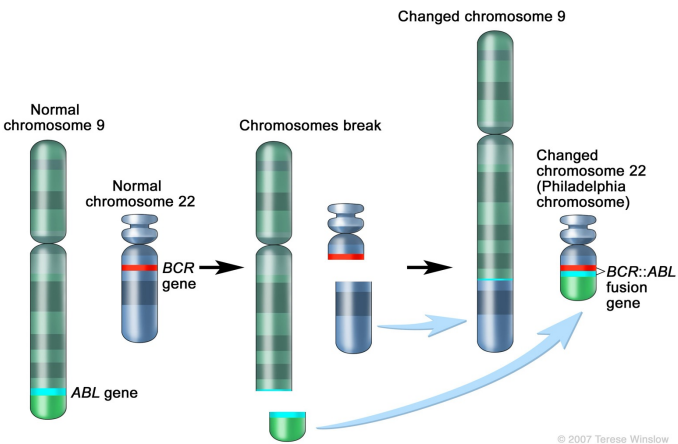
Nature volume 243, pages290–293 (1973)

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Genomic rearrangement results in oncogenic fusion gene



- Exchange of genetic material between chromosome 9 and chromosome 22 produces novel oncogenic fusion gene
 - *BCR::ABL1* creates a constitutively active tyrosine kinase



<https://www.cancer.gov/publications/dictionaries/cancer-terms/def/bcr-abl-fusion-gene>

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Understanding mechanism of *BCR::ABL1* transforms care for CML patients

UNC SCHOOL OF MEDICINE

The NEW ENGLAND JOURNAL of MEDICINE

Efficacy and Safety of a Specific Inhibitor of the BCR-ABL Tyrosine Kinase in Chronic Myeloid Leukemia

Authors: Brian J. Druker, M.D., Moshe Talpaz, M.D., Debra J. Resta, R.N., Bin Peng, Ph.D., Elisabeth Buchdunger, Ph.D., John M. Ford, M.D., Nicholas B. Lydon, Ph.D., Hagop Kantarjian, M.D., Renaud Capdeville, M.D., Sayuri Ohno-Jones, B.S., and Charles L. Sawyers, M.D. Author Info & Affiliations

- 2001 Phase I clinical trial
 - 98% had complete hematologic response
 - 54% had cytogenetic response
- Imatinib is the first FDA approved drug to counteract molecular defect = targeted therapy

Derivative chr 22 Philadelphia chr

Y = Tyrosine P = Phosphate

Mughai TI & Goldman JM. *Front Biosci.* 2006. 11:209-220.

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Combining molecular methods to diagnosis and monitor CML

UNC SCHOOL OF MEDICINE


Number of leukemic cells	Response	BCR-ABL/Control gene ratio (International Scale)
10^2	Diagnosis, pretreatment, or hematologic relapse	100
10^4	Complete hematologic response	10
10^6	Complete cytogenetic response	1
10^7	Major molecular response	0.01
10^8	Undetectable transcript (complete molecular response)	0.0001

Normal Abnormal

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Summary



- Incorporating precision medicine into oncology treatment strategies improves patient outcomes across multiple cancer types
 - More matched therapies are on the way
- Testing for genomic or molecular biomarkers can be performed on tumor tissue or via a liquid biopsy
 - Use of single gene tests versus broad, multi-gene panel options
- Increased detection of molecular biomarkers leading to approval of more targeted therapies

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Questions?

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UNC Lineberger Cancer Network **Questions/Comments?**


Nobody has responded yet.
Hang tight! Responses are coming in.

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Thank You . . .

University Cancer Research Fund

 **LINEBERGER COMPREHENSIVE CANCER CENTER**

UNC Lineberger Cancer Network

The Telehealth Team

Tim Poe – Director

Veneranda Obure – Technology Support Specialist **Andrew Dodgson, DPT** – Continuing Education Specialist

Jon Powell, PhD – Continuing Education Specialist **Patrick Muscarella** – Technology Support Technician

Oliver Marth – Technology Support Technician **Lindsey Reich, MA** – Public Communication Specialist

Barbara Walsh, DNP, MPH, MSN, RN – Nurse Planner

The song *Back Rhodes* written and performed by **Don Poe**

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Upcoming Live Webinars

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ADVANCED PRACTICE PROVIDER **Live Webinar**

May 15
4:00 PM

Using Acceptance and Commitment Therapy to Help Cancer Survivors Move Forward After Treatment

Melissa Holt, DNP, PMHNP-BC **Lisa Kanser, PsyD**

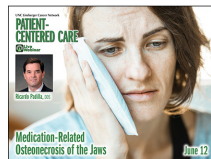


RESEARCH TO PRACTICE **Live Webinar**

May 22
12:00 PM

The Selective Use of Radiation in Solid Malignancies

Kevin Pearlstein, MD



PATIENT-CENTERED CARE **Live Webinar**

June 12
12:00 PM

Medication-Related Osteonecrosis of the Jaw

Ricardo Padilla, DDS

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ADVANCED PRACTICE PROVIDER **Self-Paced Online Course**

Physical Therapy Approaches to Oncology Care: Beyond Lymphedema

Sarah Richardson, PT, DPT, CLT, WCS



RESEARCH TO PRACTICE **Self-Paced Online Course**

Immune (check point) Related Adverse Events

Frances Collichio, MD



PATIENT-CENTERED CARE **Self-Paced Online Course**

Oncologic Emergencies

Jake Stein, MD

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